#### Astrocytes, Neurons, and Enteroviruses

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- First identified in California during the 1960s
- Infection results in "common cold-like" symptoms
- Outbreaks in the United States occurred during summers of 2014, 2016 and 2018
- Associated with "a polio-like" flaccid myelitis-626 CDC confirmed cases



### **Enterovirus D68** (EV-D68)

D68 circulation<sup>12-14</sup>



Month, year

# Virus 🛨

#### **EV-D68** pathogenesis

- Virus infection initiates in nasopharyngeal cavity
- Infection results in respiratory disease
- No respiratory epithelium damage
- instability
- Virus in CSF in 4/626 cases
- Immunoreactivity to EV-D68 VP1 peptides in CSF of 43% and sera of 73% AFM patients (Mishra et al., mBio. 2019)
- Motor cortex lesions similar to those of poliovirus

No virus isolated from blood or stool of patients, pH







Brain/meninges

#### Neurotropism, neurovirulence and neuroinvasion

- infection of cells within the central nervous system
- central nervous system
- periphery

• **Neurotropism**- the production of infectious progeny resulting from

• Neurovirulence- disease that results from infection of cells within the

• Neuroinvasion - entry into the central nervous system from the

#### **EV-D68 isolates**

- Fermon (California, 1962)
- Rhyne (California, 1962)
- New York (NYC, 2009)
- 947 (MO, 2014)
- 949 (MO, 2014)

- 952 (IL, 2014)
- 953 (KY, 2014)
- 956 (IL, 2014)
- 23087 (US, 201
- 23088 (US, 201
- 23089 (US, 201



## 0 \* \*

0.0

# Is EV-D68 neurotropic?



#### **EV-D68 replication in iCell human Neurons**



**GFAP** positive human astrocytes



#### **EV-D68 replication in human iAstrocytes**



**Hours Post Infection** 

#### EV D-68 replication in mouse postnatal brain slices

![](_page_10_Figure_1.jpeg)

![](_page_10_Figure_2.jpeg)

DAPI

Anti-enterovirus

Nissl

# EV-D68 replication in isolated glial fibrillary acidic protein (GFAP) positive mouse astrocytes

![](_page_11_Picture_1.jpeg)

from P1-P3 mice

![](_page_11_Figure_3.jpeg)

		4 0 9	NY 6
Does type I	/ml	10 <sup>8</sup>	
Interferon repress		10 <sup>7</sup>	
<b>EV-D68</b>		10° 10 <sup>5</sup>	
reproduction in	PFU	10 <sup>4</sup>	
· Sk-N-Sh cell	1 1 1	10 <sup>3</sup> 10 <sup>2</sup>	
culture?		10 <sup>1</sup>	
		10 <sup>0</sup>	

![](_page_12_Figure_2.jpeg)

4 week old C57BL/6 mice lacking the IFN α/β receptor develop paralysis after EV-D68 intracranial inoculation

![](_page_13_Figure_1.jpeg)

#### EV-D68 replication is enhanced in astrocytes isolated from IFNAR-/mice C57/B6

![](_page_14_Figure_1.jpeg)

#### Type I IFN modulates EV-D68 neurotropism

#### IFNAR<sup>-/-</sup> mice are not sufficient to study to EV-D68 associated pathologies

	1×10 <sup>8</sup> _	
Reproduction	1×10 <sup>7</sup> -	
of F\/_D68	1×10 <sup>6</sup> -	
	1×10⁵- Ε	
isolates in	1×10⁴-	
astrocytes	1×10 <sup>3</sup> -	
purified from	1×10 <sup>2</sup> -	
· outhrod mico	1×10'-	
	1×10°−	

![](_page_16_Picture_1.jpeg)

**Hours Post Infection** 

#### **10<sup>7</sup> 10**<sup>6</sup>-**10**<sup>5</sup>- $10^{4}$ Not all genetic backgrounds are 10<sup>2</sup> 10<sup>1</sup> equal **10**<sup>0</sup>-

**10**<sup>-1</sup>

![](_page_17_Figure_3.jpeg)

#### EV-D68 and poliovirus are not the same

![](_page_18_Figure_1.jpeg)

**Hours Post Infection** 

Enterovirus A71,	1×10 <sup>6</sup>
enterovirus-D68	1×10 <sup>5</sup> -
and	1×10 <sup>4</sup> -
	E 1×10 <sup>3</sup>
Coxsackievirus	<mark>ዜ</mark> 1×10²−
A16 need	1×10 <sup>1</sup> -
different genetic	1×10 <sup>0</sup> -
	1×10 <sup>-1</sup> ┘
<b>background tor</b>	2468
reproduction	、 イ

![](_page_19_Figure_1.jpeg)

**Hours Post Infection** 

## Summary

- specific
- those necessary for poliovirus, enterovirus A71 and **Coxsackievirus** 16

• EV-D68 infection is sensitive to the presence of type I Interferon • Genetic requirements for neurotropism of EV-D68 are isolate

• Genetic requirements for neurotropism of EV-D68 differ from

# Many pathways leads to neurotropism, a wrong turn for a respiratory virus

Viral infections of the CNS are less dependent on encounter with a potentially neuropathogenic agent than on some flaw in the usual barriers that normally exclude viruses from invading and infecting susceptible cells of the CNS- Richard T Johnson, 1982

## Audrey Warren

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![](_page_22_Picture_5.jpeg)

#### Animal models for studying AFM

![](_page_23_Picture_1.jpeg)

#### Kenneth Tyler, MD.

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![](_page_23_Picture_4.jpeg)